

Kinetic of Lipoquinone and Pigment Synthesis in Green *Hordeum* Seedlings during an Artificial Day-Night Rhythm with a Prolonged Dark Phase

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Chloroplast Prenyllipid Synthesis, Light-Dark Change, Thylakoid Turnover

The effect of a prolonged darkness (48 h) with a following re-illumination on the prenyllipid metabolism of chloroplasts is tested in green *Hordeum* seedlings. Continuous darkness induces thylakoid and prenyllipid breakdown and changes the remaining lipoquinone and carotenoid metabolism of chloroplasts to that of etioplasts. Re-illumination, in turn, reverses the effects of darkness and regenerates the photosynthetic apparatus within 24 hours by a continuous and specific synthesis of certain thylakoid prenyllipids.

1. The level of chlorophylls, vitamin K_1 and that of the oxidized benzoquinone forms, plastoquinone-9 and α -tocoquinone, decreases continuously in darkness. All these prenyllipids are re-synthesized upon re-illumination, whereby a faster destruction rate in the dark (α -tocoquinone + plastoquinone-9 > vitamin K_1 > chlorophylls) corresponds to a faster re-accumulation rate in the light.

2. The concentration of the reduced benzoquinones plastoquinone-9 and α -tocopherol, which are preferentially deposited in the osmiophilic plastoglobuli, increases until 24 hours of darkness and decreases thereafter either continuously (plastoquinone-9) or exhibits another strong increase (α -tocopherol). Re-illumination results only in the accumulation of the oxidized quinone forms (mainly plastoquinone-9 and little α -tocoquinone); the level of plastoquinone remains, however, almost constant with a concomitant and steady decrease in the concentration of α -tocopherol. Darkness changes the main metabolite flow in benzoquinone synthesis via the formation of α -tocopherol (a trimethyl-, phytyl-1,4-benzoquinone derivative) and light (re-illumination) via the production of plastoquinone-9 (a trimethyl, solanesyl-1,4-benzoquinone).

3. Carotenoids are enriched throughout the dark phase (mainly xanthophylls, little β -carotene) with a higher accumulation rate than after re-illumination, which yields again a higher portion of β -carotene. The correlation of prenyl chain synthesis (phytyl chain for chlorophylls and vitamin K_1 , solanesyl chain for plastoquinone-9) in the light with a reduced rate of carotenoid formation is discussed with respect to prenyl biosynthesis.

4. It is concluded that breakdown and re-synthesis of thylakoids and their prenyllipids, which are described here in a prolonged dark phase with following re-illumination, also occur during natural day-night growth of plants. The turnover of the thylakoid membranes and their lipids does, however, not get visible, since the decomposition in the night is recompensated by new synthesis during day. From the destruction rate one can calculate in a first approximation the biological half life time ($\tau^{1/2}$) which gives values in the range of 7 to 2.5 days for all thylakoid prenyllipids.

Introduction

Morphogenesis of green plants is controlled by light, which is effective *via* the photoconvertible phytochrome system¹. The active phytochrome P_{fr} also regulates growth and development of photosynthetic active chloroplasts including the formation of their photochemical active thylakoids and prenyllipids². Dark grown etiolated *Hordeum* seedlings contain thylakoid-free etioplasts whose metabolism is changed to that of chloroplasts within 24 to 30 hours of illumination with white light³. On the other hand, light cultivated green *Hordeum*

plants, when brought into continuous darkness, change their carotenoid and lipoquinone metabolism to that of etiolated plants already after a dark period of about 24 hours⁴. While chlorophylls and naphthoquinone vitamin K_1 are destroyed successively in continuous darkness, there is still an increase of the plastoquinone-9 (a dimethylbenzoquinone derivative) and β -carotene level up to 24 hours of darkness. The trimethylbenzoquinone derivative α -tocopherol and the xanthophylls, the main etioplast carotenoids, in turn, are accumulated up to three days of darkness; their concentration is decreased thereafter by destruction⁴.

In the present paper we examine whether the changes in the lipid metabolism of chloroplasts, *e. g.* immediate destruction of some prenyllipids and in-

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creased synthesis of others as induced by continuous darkness may be reversed by light. In addition, it is shown that this system of prolonged darkness with following re-illumination represents a model system for the examination of the turnover rate of thylakoids and their prenllipids.

Methods

Barley seedlings (*Hordeum vulgare* L. Asse) were cultivated in a growth chamber (24 °C, 50% relative humidity) in continuous white light (1800 lx, Osram Fluora lamps 55 W) until the age of 6 days (initial value). Thereafter all plants were brought into darkness and kept there for two days and then either re-illuminated for two days or left in darkness for two more days. The lipid analysis was performed with 200 plants each (primary leaves + coleoptile). The leaf material was homogenized and extracted at 5 °C with an acetone-petrol ether mixture in an "Ultraturrax-Homogenisator". Plastid pigments and lipoquinones were separated by thin-layer chromatography. The quantitative estimation was carried out photometrically³. The total nitrogen content was determined according to the method of Kjeldahl. The results represent mean values of several experiments.

Results

In a previous paper we have shown that six day old green *Hordeum* seedlings exhibit under exclusion of light not only the typical morphological signs of etiolement but also a change in the synthesis of their chloroplast prenllipids⁴. This change in prenllipid composition is a result of the fact that the individual thylakoid lipids are synthesized in the dark not at all (chlorophylls, vitamin K₁) or with a different rate to the light (lipophilic benzoquinones, carotenoids).

These former results are confirmed in this investigation. In addition, we have now determined the dry weight of the shoots, their total nitrogen and their total lipid content in continuous darkness. These data also show that two days of darkness are sufficient to induce in the plant catabolic processes. Light reverses these as is shown by the immediate increase of dry weight, of the lipid content and the total nitrogen level (Table I). Length growth of shoots and roots, which is promoted in darkness, slows down upon re-illumination (Fig. 1). Further-

Table I. Changes in analytical data of green 6 day old *Hordeum* seedlings in prolonged darkness and upon re-illumination (values in g per 100 shoots).

	Light 6 d	Dark +1 d	Dark +2 d	Light +1 d	Light +2 d
dryweight	1.51	1.81	1.55	1.64	1.91
total lipid content	0.175	0.170	0.143	0.166	0.193
total nitrogen content	0.055	0.068	0.066	0.069	0.074
water content	12.2	11.1	12.8	13.6	15.5

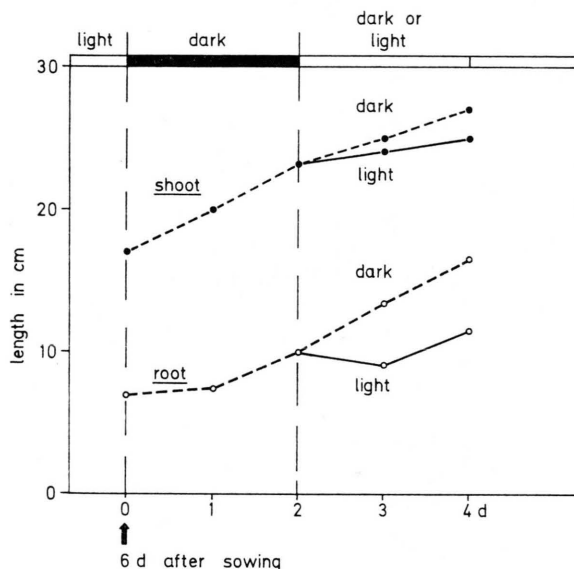


Fig. 1. Length growth of 6 day old green *Hordeum* seedlings in light-dark change.

more, light cancels out the changes in prenllipid formation induced by darkness. The synthesis rate in the light compensates and exceeds their destruction rate in darkness which results in an increase of the concentration of the major chloroplast prenllipids (Fig. 2).

a. Chlorophylls

In continuous darkness chlorophyll a and b are destroyed successively. After two days about 15% are destroyed. The chlorophyll decomposition carries on in darkness. Upon re-illumination (after two days of darkness) the chlorophyll level increases and reaches the initial value after 24 hours (Fig. 2). Destruction and new synthesis rate are about equally valid for chlorophyll a and b. Thus, the chlorophyll a/b ratio with 2.9 to 3.1 remains about the same during dark and light period.

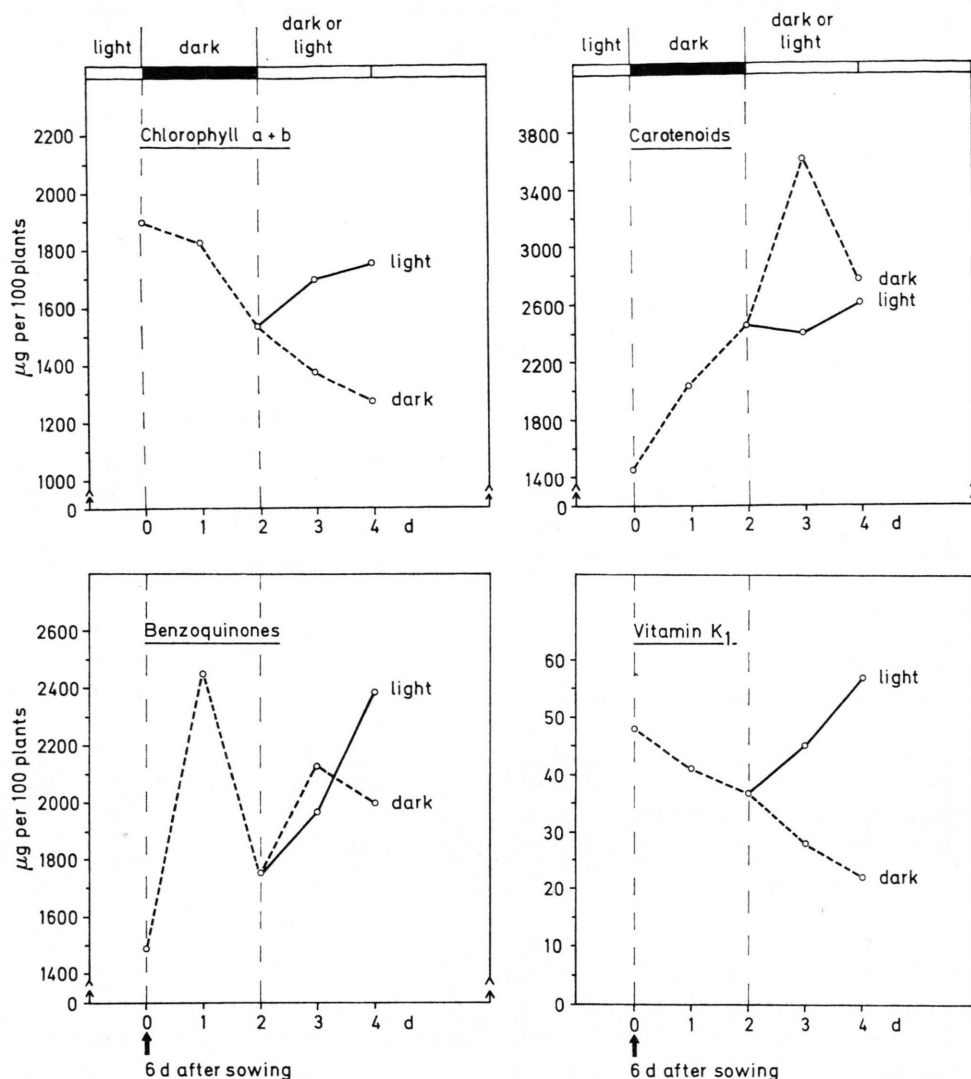


Fig. 2. Accumulation and destruction of chloroplast pigments and lipoquinones in green *Hordeum* seedlings in light or darkness.

b. Carotenoids

In contrast to the chlorophylls, the synthesis of total carotenoids continues in darkness and leads to a considerable increase of the xanthophyll content from the second to the third day of continuous darkness. Only thereafter can a decrease of the total carotenoid level by decomposition be visualized (Fig. 2). β -Carotene, whose synthesis is strongly promoted by light, is enriched in the dark only up to 24 hours. Its concentration then remains on the same level to that of the second day of darkness and decreases thereafter.

Re-illumination gives an immediate increase in the β -carotene level by new synthesis (Fig. 3). Lutein, the main xanthophyll component, and violaxanthin are initially enriched in darkness with the same synthesis rate as in the light. The large increase of the total carotenoid content from the second to third day of darkness which is due to a vast promotion of lutein and violaxanthin synthesis, is suppressed by re-illumination (Fig. 3). The pool-size of zeaxanthin, which is particularly enriched from the first to the second day of darkness⁴, decreases in continued darkness and also upon re-illumination. In both cases it may get transferred to

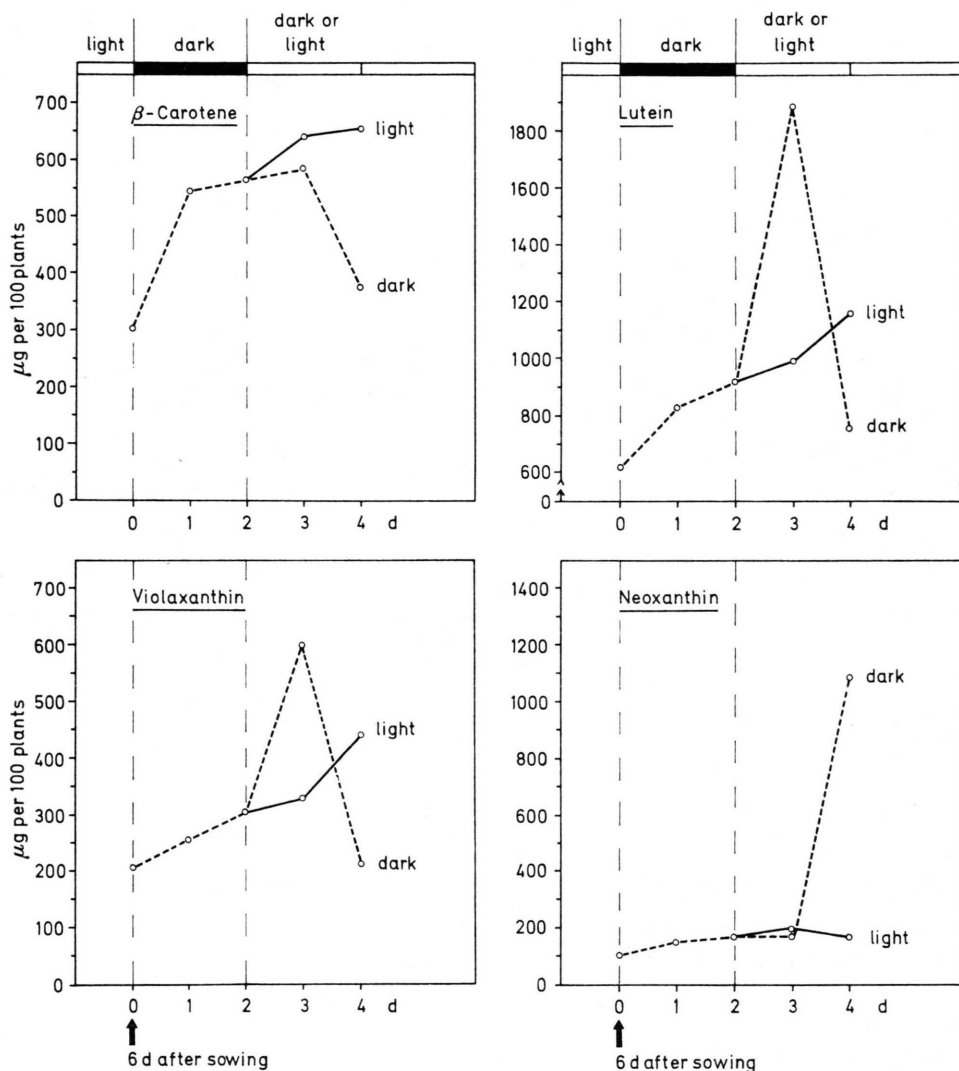


Fig. 3. Changes in the level of the main carotenoids during a prolonged dark period and upon re-illumination.

violaxanthin or to its stereo-isomere lutein. The level of the other xanthophylls, which are present in lower concentrations, are less influenced by the dark-light change.

The different synthesis rate of the single carotenoids in darkness results in a change of the percentage composition of the carotenoids. The relative portion of β -carotene, violaxanthin and lutein is diminished within two days of darkness, while the portion of the other carotenoids is increased (Table II). Re-illumination reverses this process. The results thus show that the carotenoid metabolism of chloroplasts is changed in continuous darkness successively to that of etioplasts. Upon

re-illumination the carotenoid composition of functioning chloroplasts is reached within one to two days. After re-illumination the portion of zeaxanthin with 0.8% remains significantly beyond the initial value (9% in six day old plants). This decrease in the relative zeaxanthin level, however, is dependent on the development stage and is found in the light-grown green control plants, too.

c. Plastid-quinones

Continuous darkness leads in the first 24 hours to a strong increase of the total benzoquinone content of chloroplasts. Then its level is decreased considerably by decomposition. After the second to

Table II. Changes in the percentage composition (weight %) of carotenoids in green *Hordeum* seedlings in darkness and upon re-illumination.

	Light 6 d	Dark +1 d	Dark +2 d		Dark or Light +1 d	Dark or Light +2 d	
β -carotene	21.2	27.0	23.0	\nearrow	16.2	14.7	dark
				\searrow	26.8	25.1	light
lutein	43.7	40.9	37.4	\nearrow	52.5	29.8	dark
				\searrow	41.4	44.6	light
violaxanthin	14.4	12.7	12.5	\nearrow	16.6	8.3	dark
				\searrow	13.9	16.9	light
neoxanthin	7.4	7.7	7.0	\nearrow	4.9	42.9	dark
				\searrow	8.1	6.6	light
zeaxanthin	9.0	7.4	14.6	\nearrow	5.0	1.1	dark
				\searrow	1.9	0.8	light
luteinopoxid	0.5	0.8	1.0	\nearrow	0.8	1.3	dark
				\searrow	1.3	1.1	light
antheraxanthin	2.1	1.7	1.8	\nearrow	2.0	0.4	dark
				\searrow	3.6	2.4	light
neoxanthin a	1.7	1.8	2.7	\nearrow	2.0	1.5	dark
				\searrow	3.0	2.5	light
carotenoids	100.0	100.0	100.0	\nearrow	100.0	100.0	dark
				\searrow	100.0	100.0	light

fourth day of darkness there is again a low net increase of the total benzoquinone concentration (Fig. 2). The individual benzoquinones, which consist of the two lipophilic redox systems plastoquinone-9 (+ plastohydroquinone-9) and α -tocopherol (+ α -tocoquinone) behave, however, in a different way.

The level of total plastoquinone-9 is increased only up to 24 hours of darkness and decreases thereafter successively (Fig. 4). The concentration of the oxidized form (plastoquinone-9) decreases immediately in darkness.

Re-illumination results in an increase of total plastoquinone-9 content which is due exclusively to the formation of the oxidized form plastoquinone-9. The level of plastohydroquinone-9, in turn, is decreased in the light in a similar way as in continued darkness (Fig. 4).

The level of the trimethyl-, phytyl-1,4-benzoquinone derivative α -tocopherol shows an increase up to 24 hours of darkness and a significant decrease thereafter. Further darkness results in a considerable re-accumulation of α -tocopherol, which parallels in time the decomposition of chlorophylls and which is similarly found during chloroplast degeneration⁶ and during development of chromoplasts from chloroplasts^{7,8}. The level of its oxidized form α -tocoquinone, however, decreases successively in continuous darkness (Fig. 4). Upon re-illumina-

tion the concentration of α -tocopherol decreases continuously in the light. The level of its oxidized form α -tocoquinone, which is present in much lower concentration, however, is raised in the light. In contrast to the benzoquinones the synthesis of the naphthoquinone vitamin K₁ is similarly regulated to that of the chlorophylls (Fig. 2). The K₁ concentration decreases continuously in darkness and is raised again upon re-illumination. These light and dark changes in benzoquinone and vitamin K₁ synthesis result in the differences in the percentage composition of the lipoquinones as is shown in Table III. The relative portion of α -tocopherol in-

Table III. Percentage composition (weight %) of chloroplast lipoquinones in 6 day old green barley seedlings in a prolonged dark phase and upon re-illumination.

	6 d	Dark +1 d	Dark +2 d		Dark or Light +1 d	Dark or Light +2 d	
plasto-quinone-9	17.5	6.9	7.4	\nearrow	4.0	2.0	dark
				\searrow	17.9	40.8	light
plastohydro-quinone-9	43.1	50.1	49.9	\nearrow	42.6	41.4	dark
				\searrow	44.8	31.5	light
α -tocopherol	34.3	40.7	40.4	\nearrow	51.9	55.4	dark
				\searrow	34.8	24.3	light
α -tocoquinone	2.0	0.7	0.2	\nearrow	0.1	0.1	dark
				\searrow	0.2	1.1	light
vitamin K ₁	3.1	1.6	2.1	\nearrow	1.4	1.1	dark
				\searrow	2.3	2.3	light
total lipo-quinones	100.0	100.0	100.0	\nearrow	100.0	100.0	dark
				\searrow	100.0	100.0	light

creases with continuous darkness from 35 to 55% of total lipoquinone content while that of total plastoquinone-9 is diminished from 60 to 43% after 4 days darkness. Re-illumination reverses this process. After two days light the total plastoquinone-9 content rises up to 72%, while that of α -tocopherol decreases to 24% of total chloroplast lipoquinones. The relative lipoquinone composition in darkness thus resembles that of etioplasts and upon re-illumination again that of chloroplasts.

d. Prenyl synthesis

Lipoquinones and chlorophylls with their isoprenoid side chains belong, together with the carotenoids, to the group of isoprenoid plastid lipids (prenyllipids). Phytosterols also belong to this group, but they are present in chloroplasts only in traces. Thus by formation of the sum of carotenoids and prenyl side chains one obtains a rough approximation on the extent of total chloroplast prenyl syn-

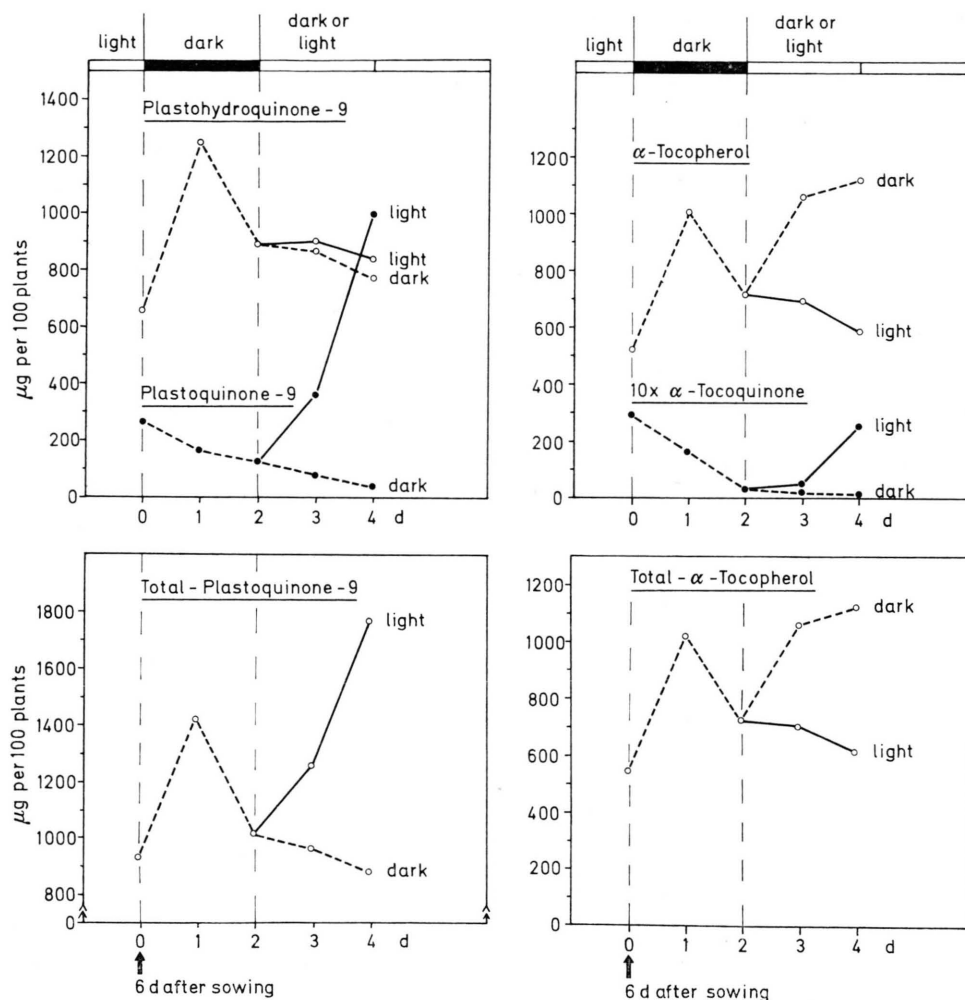


Fig. 4. Changes in the benzoquinone content of green *Hordeum* seedlings in darkness and upon re-illumination.

Table IV. Increase and decrease in the level of C_{20} -, C_{40} - and C_{45} -prenyl-chains in green 6 day old *Hordeum* seedlings in a prolonged dark period and upon re-illumination.

	Light 6 d	+1 d	Dark +2 d		+1 d	Dark or Light +2 d	
sum of C_{20} chains	981.2	1263.7	970.5	↗	1141.6	1140.3	dark
				↘	1012.1	988.1	light
C_{20} in chlorophylls	591.2	568.5	476.4	↗	427.8	395.3	dark
				↘	526.7	548.8	light
C_{20} in α -tocopherol (+ α -tocoquinone)	360.2	669.8	471.2	↗	696.4	731.4	dark
				↘	457.5	403.0	light
C_{20} in vitamin K_1	29.8	25.4	22.9	↗	17.4	13.6	dark
				↘	27.9	35.3	light
C_{45} chains (plastoquinone-9 + hydroquinone)	765.6	1173.7	844.0	↗	793.7	726.0	dark
				↘	1039.5	1460.3	light
C_{40} chains (carotenoids)	1422.0	2023.5	2456.0	↗	3603.0	2540.0	dark
				↘	2388.0	2602.0	light
sum of prenyl-chains	3168.8	4459.9	4270.5	↗	5538.3	4406.3	dark
				↘	4439.6	5048.4	light

thesis. It should be considered in this respect that only the side chains bound to prenallipids are determined in this investigation but not free side chains which may originate, *e.g.* from the decomposition of chlorophylls.

The total prenall chain content of chloroplasts increases up to the first day of darkness parallel to the dark formation of plastohydroquinone-9, α -tocopherol and carotenoids (Table IV). There is a small decrease in total prenall content from the first to the second day of darkness, which is a result of the decomposition of chlorophylls, vitamin K_1 , plastohydroquinone-9, α -tocopherol and α -tocoquinone, and which does not get compensated by the simultaneous accumulation of carotenoids. With the strongly increased formation of carotenoids from the second to the third day of darkness which parallels the re-accumulation of α -tocopherol there is a considerable increase in the total prenall content. The latter decreases thereafter by decomposition. Re-illumination, on the other hand, leads as expected, to a continuous augmentation of the prenall content, which parallels the light induced re-accumulation of chlorophylls, vitamin K_1 , plastohydroquinone-9 and α -tocoquinone. The individual side chains, however, show a different behaviour. The level of the C_{45} -chain declines continuously after the first day of darkness and is increased upon re-illumination. The phytol chain concentration shows after initial augmentation, which parallels the increase in the α -tocopherol level (first day of darkness), a marked decline on the second day, and increases once more to a constant level in further darkness. Re-illumination does not influence total phytol chain content since the increase in chlorophyll, K_1 and α -tocoquinone level is compensated in this respect by the decomposition of α -tocopherol.

Discussion

The results of our investigation with a prolonged dark phase followed by re-illumination shows that the total pigment and lipoquinone metabolism of chloroplasts is regulated by light. Darkness, which inhibits the formation of thylakoids, changes the relative prenallipid concentrations of chloroplasts to that of etioplasts. The concentration of chlorophylls, vitamin K_1 and of the oxidized form of the benzoquinone derivatives (plastohydroquinone-9, α -tocoquinone) decreases immediately in darkness. The

destruction of these prenallipids is strong indication for thylakoid breakdown. The reduced benzoquinone forms α -tocopherol and plastohydroquinone in turn, which are located only to a small extent in thylakoids and get mainly deposited in the osmophilic plastoglobuli^{9,10}, are strongly enriched in the first 24 hours of darkness. This is similar to the natural breakdown of thylakoids. In contrast to natural chloroplast breakdown, where carotenoids get decomposed in a kinetic similar to that of chlorophylls⁶, they are enriched up to three days of darkness with a predominant formation of xanthophylls and less β -carotene. Thus, as seen from the prenallipid analysis, the chloroplasts of *Hordeum* seedlings show after two days of darkness all signs of thylakoid breakdown whereby the remaining capacity for prenallipid synthesis corresponds to that of etioplasts.

Re-illumination of the *Hordeum* seedlings, which were kept two days in darkness, regenerates the photosynthetic apparatus by new formation of thylakoids. The accumulation of chlorophylls, vitamin K_1 , of solely oxidized benzoquinones (plastohydroquinone-9, α -tocoquinone) and of a higher portion of β -carotene are indication for active thylakoid formation. The relative chloroplast lipid concentrations of the darkened seedlings are reconverted to that of intact chloroplasts within 24 hours of re-illumination. The kinetic of prenallipid formation upon re-illumination resembles very much that during the first greening of dark grown, etiolated seedlings³.

The chromanol α -tocopherol, which becomes accumulated in continued darkness at high rates, is not synthesized upon re-illumination. From this one may conclude that α -tocopherol has no primary function in photosynthetic processes and plays only a role as potential lipid antioxidant¹¹ in the preservation of the photosynthetic biomembrane. The level of plastohydroquinone-9 (oxidized form), the terminal electron acceptor of the photosynthetic light reaction II which also promotes cyclic electron flow¹², in turn is raised considerably upon re-illumination. In view of the well established biosynthesis route of isoprenoid benzoquinones^{8,16}, our observations thus indicate that the common benzoquinone precursor, homogentisic acid, is bound in the light mainly to the C_{45} -chain to form plastohydroquinone-9, and in darkness predominantly to the phytol chain to yield α -tocopherol. This light

induced preferential synthesis of plastoquinone-9 is always found during active thylakoid formation and is further enhanced by the phytohormones kinetin¹³ and β -indolacetic-acid (IAA)¹⁴, while darkness and gibberellic acid (GA₃)¹³ result in a preferential α -tocopherol accumulation.

Vitamin K₁ and α -tocoquinone, which are present in chloroplasts in minor concentration, are accumulated upon re-illumination in a similar pattern as the chlorophylls. This makes their participation as electron carriers in photosynthetic reactions very probable, though their exact position in the photosynthetic electron transport chain is not yet known. Vitamin K₁ which is bound within the chloroplasts to the photosystem I particles¹⁵ is a possible candidate for the unknown endogenous promoter of cyclic electron flow.

Between light and darkness there is also a particular difference in the formation of the individual chloroplast prenyl chains. Though the level of chlorophylls and vitamin K₁ is decreased continuously in darkness, there seems to occur phytyl chain formation in dark as is indicated by the dark accumulation of α -tocopherol, in which the phytyl chain is built in, too (Fig. 5). Since it can not be ex-

prenyl chain (solanesyl-chain of plastoquinone-9) apparently ceases after 24 hours of darkness. The formation of geranyl-geranyl-pyrophosphate, as common precursor of all chloroplast prenyl chains, and its dimerisation to the tetraterpenoid chain of carotenoids continues, however, in the dark. Re-illumination in turn gives more phytyl chains (chlorophylls, vitamin K₁), induces again the formation of the solanesyl chain (plastoquinone-9) and reduces the rate of carotenoid formation. It has also been shown in some other cases that enhanced chlorophyll synthesis is accompanied by a decrease in the accumulation rate of carotenoids¹³.

The results of this investigation show that all chloroplast prenyllipids tested here undergo destruction. Thus the chloroplasts must possess the various enzymes for breakdown and metabolisation of such different prenyllipids as naphtho- and benzoquinones as well as chlorophylls and carotenoids. The destruction of these chloroplast lipids is, however, only visualized in a prolonged dark phase, when the light induced new formation of chloroplast prenyllipids is cancelled out. We thus conclude from our results with the prolonged dark phase that turnover of thylakoid prenyllipids and possibly all other thylakoid components must exist in chloroplasts of young and adult leaves.

From the decrease rate of the prenyllipids in continuous darkness one can calculate in a rough approximation their turnover rate. For the chlorophylls a and b one obtains a turnover rate of about 8% per day which corresponds to a biological half life time $\tau^{1/2}$ of six days. It is of interest in this respect that the turnover rate for chlorophylls of other plants, as estimated from isotopic studies, were found in the range of 5 to maximal 10% per day¹⁷. Vitamin K₁ seems to have a much higher turnover rate of about 13% per day ($\tau^{1/2}$ = 4 days). Its rate of re-accumulation upon re-illumination of the darkened seedlings is also faster than that of chlorophylls. For the oxidized benzoquinone forms plastoquinone-9 and α -tocoquinone, which are mainly localized in the thylakoids, the calculation yields turnover rates of about 20% per day ($\tau^{1/2}$ = 2.5 days). The reduced benzoquinone forms (α -tocopherol and plastoquinone-9), which are preferentially deposited in the osmiophilic plastoglobuli of the chloroplast stroma⁹, seem to have lower turnover rates than their oxidized forms. Since both reduced benzoquinone derivatives exhibit an initial

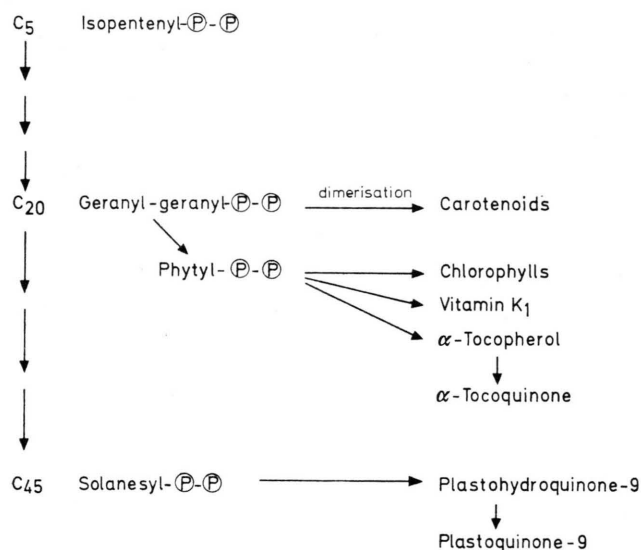


Fig. 5. Scheme of prenyl-biosynthesis (isoprenoid side chains and carotenoids)⁸.

cluded that some phytyl chains, which may derive from chlorophyll breakdown, are used for α -tocopherol formation, the synthesis rate of the phytyl chain in darkness appears to be, however, much lower than in the light. The synthesis of the C₄₅-

fast destruction, which then slows down, one can calculate two turnover rates. These are for plasto-hydroquinone-9, 7 or 13% per day ($\tau^{1/2} = 7$ or 4 days) and for α -tocopherol 8.5 or 13% per day ($\tau^{1/2} = 6$ or 4 days). Since carotenoids are accumulated even in darkness and in the light it is not possible from our results to estimate their destruction rate.

In any case, this rough approximation shows that the different thylakoid prenyllipids exhibit individual half life time ($\tau^{1/2}$) which amount to six days or less, and which may be similar for other

thylakoid components. It is of particular interest that a faster destruction rate corresponds to a faster rate of re-accumulation of the individual prenyllipid in the light. This shows the usefulness of this system with prolonged dark phase and re-illumination for the investigation of the turnover of thylakoids and their lipid and protein components. Further experiments to determine turnover rates of thylakoid lipids with ^{14}C labelling are in progress.

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